

Perspectives on Biofilm Challenges and Solutions

Ravindra Wavhale

Department of Pharmaceutical Chemistry, Dr. D. Y. Patil Institute of Pharmaceutical Sciences and Research, Pune, Maharashtra, India

ABSTRACT

Biofilms consist of an extracellular polymeric matrix that protects microorganisms from external stresses, such as antibiotics and the host immune response, making microbial infections difficult to treat. Biofilm-associated infections are on the rise, often leading to chronic conditions and resistance to multiple drugs, resulting in increased mortality rates. This necessitates special consideration when selecting materials for implants, lenses, sutures, prosthetics, and other medical devices. In this review, we discuss various approaches for biofilm inhibition, their mechanisms of action, and the challenges associated with their clinical translation.

Keywords: Antibiofilm agents, bacterial infection, biofilm, drug resistance, implants


Biofilms are formed when microorganisms adhere to one another and solid surfaces. These attached cells get embedded in a matrix made up of extracellular polymeric substances, creating a thin film that provides a protected environment for the microorganisms.^[1-3] Approximately 80% of infections are associated with biofilm formation,^[4] which is commonly found on medical devices^[5] such as knee replacements, catheters, implants, prosthetic valves, contact lenses, screws, pins, and joints. This biofilm formation significantly affects antimicrobial efficacy and immune responses,^[6] contributing substantially to antimicrobial resistance^[7,8] and presenting serious healthcare challenges. To address this issue, various strategies have been implemented, including the use of antimicrobial peptides,^[9-11] modifications to medical devices,^[12,13] quaternary ammonium compounds,^[14-16] compounds

that release nitric oxide,^[17,18] inhibitors of cell signaling pathways,^[19-22] antibiotic conjugates,^[23-25] photothermal therapy (PTT),^[26] and surgical removal of biofilm biomass.^[27]

Antimicrobial peptides, composed of 10–50 amino acids, exhibit potent antimicrobial and antibiofilm activity. Their antimicrobial mechanism involves permeating cell membranes, leading to the formation of membrane-spanning pores. This destabilizes the membrane and neutralizes or disaggregates lipopolysaccharides. However, their antibiofilm activity is attributed to their ability to disrupt cell membranes and interfere with quorum sensing, thereby inhibiting the adhesion of bacterial cells to solid surfaces.

Another promising strategy for preventing microbial biofilm formation on invasive medical devices involves modifying their surfaces using biocides or developing anti-adhesive materials. However, there is ongoing debate regarding biocide coatings, particularly concerning the potential development of antibiotic resistance due to long-term use and the release of sub-inhibitory drug concentrations.

Quaternary ammonium salts (QAS) have also been proven to demonstrate antibiofilm activity by penetrating the cell wall and interacting with lipids and proteins in the cell membrane, leading to structural

Access this article online	
Website: themmj.in	Quick Response Code 
DOI: 10.15713/ins.mmj.109	

Address for correspondence:

Ravindra Wavhale, Department of Pharmaceutical Chemistry, Dr. D. Y. Patil Institute of Pharmaceutical Sciences and Research, Pune, Maharashtra, India. E-mail: ravindra.wavhale@dypvp.edu.in

disorganization and ultimately causing cell lysis. The activity and selectivity of QAS depend on their chain length, charge, and monomeric or dimeric nature.

An alternative strategy for biofilm disruption is the use of exogenous nitric oxide (NO)-releasing agents. This approach presents a promising option due to its ability to disperse and inhibit biofilms. NO donors, including small molecules, macromolecules, and nanoparticles, can be utilized to generate highly reactive NO, which interacts with oxygen, transition metals, DNA, and proteins, resulting in potent antibiofilm and antimicrobial effects.

Molecular-level studies on the mechanism of biofilm formation have revealed that cyclic-di-GMP, a secondary signaling molecule, plays a crucial role in the biofilm life cycle. High levels of cyclic-di-GMP are directly linked to biofilm formation, whereas reduced levels promote biofilm dispersal. Further research has demonstrated that diguanylate cyclase (DGC) catalyzes the synthesis of cyclic-di-GMP, while phosphodiesterase (PDE) facilitates its degradation. Consequently, several DGC and PDE modulators have been identified to regulate cyclic-di-GMP levels, offering a potential strategy to control biofilm formation.

The development of resistance to conventional antibiotics is largely attributed to reduced drug permeation through the exopolymeric matrix of biofilms. To overcome this resistance mechanism, a promising approach involves the use of antibiotic conjugates, where antibiotics are linked to membrane-penetrating peptides or polymers. In addition, conjugating antibiotics with nanoparticles offers a valuable strategy for enhancing drug penetration through bacterial biofilms.

Another promising approach for combating biofilms is PTT, which offers several advantages, including its non-invasive nature, broad-spectrum activity, shorter treatment duration, and low systemic toxicity. The effectiveness of PTT depends on laser properties and the selection of suitable photothermal agents. Furthermore, PTT has shown significant potential as a combination therapy when used alongside antimicrobial agents and nanotechnology, enhancing its overall efficacy against biofilms.

Over the past decade, research has increasingly focused on designing and developing nano/micromotors (NMMs) to effectively disrupt microbial biofilms.^[28-36] NMMs are notable for their ability to harness external or environmental energy and convert it into propulsion. This capability can be strategically employed for the physical and chemical disruption

of biofilms. The mechanism involves the deeper penetration of NMM into the extracellular polymeric matrix by utilizing driving forces from external or internal sources, such as hydrogen peroxide, magnetic fields, or ultrasound. Although NMM appears to be a promising strategy against biofilms and addresses serious issues of drug resistance in clinical settings, there are still future challenges to consider. These challenges include large-scale production, stability, treatment efficiency, and *in vivo* translation, all of which need to be addressed in future research in this area.

Biofilms are now recognized as a critical global issue impacting the healthcare, food, water, and marine industries, resulting in significant health and economic burdens worldwide. Fundamental challenges in biofilm research include the development of *in vitro* assay protocols and computer-aided models that accurately replicate real-life environments, as well as bridging the gap between oversimplified models and the highly complex and diverse nature of biofilm development. In addition, the lack of standardized guidelines for diagnosing and categorizing biofilms, testing antibiofilm agents, and providing physicians with treatment protocols for biofilm infections continues to hinder progress in biofilm research. The development of Food and Drug Administration -approved antibiofilm drugs and the translation of nanotechnology for their successful clinical applications, particularly in chronic and multidrug-resistant biofilm infections, remain major current challenges for researchers in this field.

REFERENCES

1. Donlan RM. Biofilms: Microbial life on surfaces. *Emerg Infect Dis* 2002;8:881-90.
2. Garrett TR, Bhakoo M, Zhang Z. Bacterial adhesion and biofilms on surfaces. *Prog Nat Sci* 2008;18:1049-56.
3. Donlan RM, Costerton JW. Biofilms: Survival mechanisms of clinically relevant microorganisms. *Clin Microbiol Rev* 2002;15:167-93.
4. Jamal M, Ahmad W, Andleeb S, Jalil F, Imran M, Nawaz MA, *et al.* Bacterial biofilm and associated infections. *J Chin Med Assoc* 2018;81:7-11.
5. Von Eiff C, Jansen B, Kohnen W, Becker K. Infections associated with medical devices: Pathogenesis, management and prophylaxis. *Drugs* 2005;65:179-214.
6. Rybtke M, Hultqvist LD, Givskov M, Tolker-Nielsen T. *Pseudomonas aeruginosa* biofilm infections: Community structure, antimicrobial tolerance and immune response. *J Mol Biol* 2015;427:3628-45.

7. Bowler P, Murphy C, Wolcott R. Biofilm exacerbates antibiotic resistance: Is this a current oversight in antimicrobial stewardship? *Antimicrob Resist Infect Control* 2020;9:162.
8. Dufour D, Leung V, Lévesque CM. Bacterial biofilm: Structure, function, and antimicrobial resistance. *Endod Top* 2010;22:2-16.
9. Niu Y, Yan Z, Yang S, Wang Y, Wang X, Li C, *et al.* Cholesteryl succinate-modified antimicrobial peptides for selective sterilization of *Staphylococcus aureus* in photodynamic therapy. *ACS Appl Nano Mater* 2025;8:2421-31.
10. Kumar SD, Park J, Radhakrishnan NK, Aryal YP, Jeong GH, Pyo IH, *et al.* Novel leech antimicrobial peptides, hirunipins: Real-time 3D monitoring of antimicrobial and antibiofilm mechanisms using optical diffraction tomography. *Adv Sci (Weinh)* 2025;12:e2409803.
11. Lamba S, Wang K, Lu J, Phillips AR, Swift S, Sarojini V. Polydopamine-mediated antimicrobial lipopeptide surface coating for medical devices. *ACS Appl Bio Mater* 2024;7:7574-84.
12. Francis AL, Namasivayam SK, Samrat K. Potential of silver nanoparticles synthesized from *Justicia adhatoda* metabolites for inhibiting biofilm on urinary catheters. *Microb Pathog* 2024;196:106957.
13. Kirla H, Hamzah J, Jiang ZT, Henry DJ. Dual-action antimicrobial surface coatings: Methylene blue and quaternary ammonium cation conjugated silica nanoparticles. *RSC Pharm* 2025;2:163-77.
14. Muzychka L, Hodyna D, Metelytsia L, Smolii O. Nature-inspired novel quaternary ammonium compounds: Synthesis, antibacterial and antibiofilm activity. *ChemMedChem* 2025;20:e202400807.
15. Saverina EA, Frolov NA, Kamanina OA, Arlyapov VA, Vereshchagin AN, Ananikov VP. From antibacterial to antibiofilm targeting: An emerging paradigm shift in the development of quaternary ammonium compounds (QACs). *ACS Infect Dis* 2023;9:394-422.
16. Jennings MC, Minbiole KP, Wuest WM. Quaternary ammonium compounds: An antimicrobial mainstay and platform for innovation to address bacterial resistance. *ACS Infect Dis* 2015;1:288-303.
17. Chiarelli LR, Degiacomi G, Egorova A, Makarov V, Pasca MR. Nitric oxide-releasing compounds for the treatment of lung infections. *Drug Discov Today* 2021;26:542-50.
18. Xu LC, Wo Y, Meyerhoff ME, Siedlecki CA. Inhibition of bacterial adhesion and biofilm formation by dual functional textured and nitric oxide releasing surfaces. *Acta Biomater* 2017;51:53-65.
19. Ha DG, O'Toole GA. c-di-GMP and its effects on biofilm formation and dispersion: A *Pseudomonas aeruginosa* review. *Microbiol Spectr* 2015;3:MB-0003-2014.
20. Park S, Sauer K. Controlling biofilm development through cyclic di-GMP signaling. *Adv Exp Med Biol* 2022;1386:69-94.
21. Valentini M, Filloux A. Multiple roles of c-di-GMP signaling in bacterial pathogenesis. *Annu Rev Microbiol* 2019;73:387-406.
22. Hengge R, Gründling A, Jenal U, Ryan R, Yildiz F. Bacterial signal transduction by cyclic di-GMP and other nucleotide second messengers. *J Bacteriol* 2016;198:15-26.
23. Meiers J, Zahorska E, Röhrig T, Hauck D, Wagner S, Titz A. Directing drugs to bugs: Antibiotic-carbohydrate conjugates targeting biofilm-associated lectins of *Pseudomonas aeruginosa*. *J Med Chem* 2020;63:11707-24.
24. Tvilum A, Johansen MI, Glud LN, Ivarsen DM, Khamas AB, Carmali S, *et al.* Antibody-drug conjugates to treat bacterial biofilms via targeting and extracellular drug release. *Adv Sci (Weinh)* 2023;10:e2301340.
25. Kasza K, Richards B, Jones S, Romero M, Robertson SN, Hardie KR, *et al.* Ciprofloxacin poly(β -amino ester) conjugates enhance antibiofilm activity and slow the development of resistance. *ACS Appl Mater Interfaces* 2024;16:5412-25.
26. Yuan Z, Lin C, He Y, Tao B, Chen M, Zhang J, *et al.* Near-infrared light-triggered nitric-oxide-enhanced photodynamic therapy and low-temperature photothermal therapy for biofilm elimination. *ACS Nano* 2020;14:3546-62.
27. Wang S, Zhao Y, Breslawec AP, Liang T, Deng Z, Kuperman LL, *et al.* Strategy to combat biofilms: A focus on biofilm dispersal enzymes. *NPJ Biofilms Microbiomes* 2023;9:63.
28. Mayorga-Martinez CC, Zhang L, Pumera M. Chemical multiscale robotics for bacterial biofilm treatment. *Chem Soc Rev* 2024;53:2284-99.
29. Xie S, Huang K, Peng J, Liu Y, Cao W, Zhang D, *et al.* Self-propelling nanomotors integrated with biofilm microenvironment-activated NO release to accelerate healing of bacteria-infected diabetic wounds. *Adv Healthc Mater* 2022;11:e2201323.
30. Zhong W, Handschuh-Wang S, Uthappa UT, Shen J, Qiu M, Du S, *et al.* Miniature robots for battling bacterial infection. *ACS Nano* 2024;18:32335-63.
31. Maric T, Løvind A, Zhang Z, Geng J, Boisen A. Near-infrared light-driven mesoporous SiO₂/Au nanomotors for eradication of *Pseudomonas aeruginosa* biofilm. *Adv Healthc Mater* 2023;12:e2203018.
32. Villa K, Sopha H, Zelenka J, Motola M, Dekanovsky L, Beketova DC, *et al.* Enzyme-photocatalyst tandem microrobot powered by urea for *Escherichia coli* biofilm eradication. *Small* 2022;18:e2106612.
33. Zheng J, Liu H, Deng Y, Lian L, Hua N, Zhao S, *et al.* Biofilm microenvironment-mediated dual-gases-driven nanomotors for combating drug-resistant bacterial infections. *ACS Mater Lett* 2024;6:5128-37.
34. Ziemyte M, Escudero A, Díez P, Ferrer MD, Murguía JR, Martí-Centelles V, *et al.* Ficin-cyclodextrin-

- based docking nanoarchitectonics of self-propelled nanomotors for bacterial biofilm eradication. *Chem Mater* 2023;35:4412-26.
35. Zheng J, Wang W, Gao X, Zhao S, Chen W, Li J, *et al.* Cascade catalytically released nitric oxide-driven nanomotor with enhanced penetration for antibiofilm. *Small* 2022;18:e2205252.
36. Yuan K, Jurado-Sánchez B, Escarpa A. Dual-propelled lanibiotic based janus micromotors for selective

inactivation of bacterial biofilms. *Angew Chem Int Ed Engl* 2021;60:4915-24.

How to cite: Wavhale R. Perspectives on Biofilm Challenges and Solutions. *MIMER Med J* 2025;9(1):18-21.

Source of Support: Nil. **Conflicts of Interest:** None declared.

© The Author(s). 2025 Open Access. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<https://creativecommons.org/licenses/by-nc/4.0/>), which permits unrestricted use, distribution, and non-commercial reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated.